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APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody

Catalog # ATB10421

Specification

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - Product Information

Application	**FC**
Isotype	**Armenian Hamster IgG**
Concentration	**0.2mg/ml**
Reactivity	**Mouse**
Formulation	**10 mM NaH2PO4, 150 mM NaCl, 0.09% NaN3, 0.1% gelatin, pH7.2**

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - Additional Information

Gene ID	**12519**
Gene Name	**Cd80**

Alternative Name(s)
B7, Ly-53

Format
APC

Storage Conditions
2-8°C protected from light

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - Protocols

Provided below are standard protocols that you may find useful for product applications.

- Western Blot
- Blocking Peptides
- Dot Blot
- Immunohistochemistry
- Immunofluorescence
- Immunoprecipitation
- Flow Cytomety
- Cell Culture

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - Images

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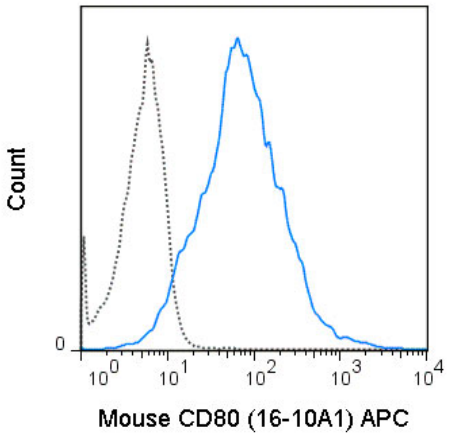

C57Bl/6 splenocytes were stimulated with anti-IgM and anti-CD40 for 4 days. Cells were then stained with 0.06 ug APC Anti-Mouse CD80 (ATB10421) (solid line) or 0.06 ug APC Armenian Hamster isotype control (dashed line).

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - Background

The 16-10A1 antibody reacts with mouse CD80, also known as B7-1, a 55 kDa type I transmembrane protein ligand for CD152 (CTLA-4) and for CD28, a co-stimulatory receptor for the T cell receptor (TCR). CD28 also binds a second B7 ligand known as CD86 (B7-2). Both CD80 and CD86 are expressed on activated B cells and antigen-presenting cells. These ligands trigger CD28 signaling in concert with TCR activation to drive T cell proliferation, induce high-level expression of IL-2, impart resistance to apoptosis, and enhance T cell cytotoxicity. The interaction / co-stimulatory signaling between the B7 ligands and CD28 or CTLA-4 provides crucial communication between T cells and B cells or APCs to coordinate the adaptive immune response.

APC Anti-Mouse CD80 (B7-1) (16-10A1) Antibody - References

Thaventhiran JED, Hoffmann A, Magiera L, de la Roche M, Lingel H, Brunner-Weinzierl M, and Fearon DT. 2012. Proc. Natl. Acad. Sci. 10.1073. (in vitro blocking, Flow cytometry)

Liu Z, Geboes K, Hellings P, Maerten P, Heremans H, Vandenberghe P, Boon L, van Kooten P, Rutgeerts P, and Ceuppens JL. 2011. J. Immunol. 167: 1830-1838. (in vivo blocking, Immunohistochemistry – OCT embedded frozen tissue)

Anraku M, Tagawa T, Wu Licun, Yun Z, Keshavjee S, Zhang L, Johnston MR, and de Perrot M. 2010. J. Immunol. 185:956-966. (Flow cytometry)

Odobasic D, Kitching AR, Semple TJ, Timoshanko JR, Tipping PG, and Holdsworth SR. 2005. J. Am. Soc. Nephrol. 16: 2012-2022. (in vivo activation, Immunofluorescence microscopy and Immunohistochemistry – frozen tissue)

Lenschow DJ, Ho SC, Sattar H, Rhee L, Gray G, Nabavi N, Herold KC, and Bluestone JA. 1995. J. Exp. Med. 181:1145-155. (in vitro blocking)

Razi-Wold Z, Freeman GJ, Galvin F, Benacerraf B, Nadler L, and Reiser H. 1992. Proc. Natl. Acad. Sci. 89:4210-4214. (Origination of clone, Immunoprecipitation, in vitro blocking)